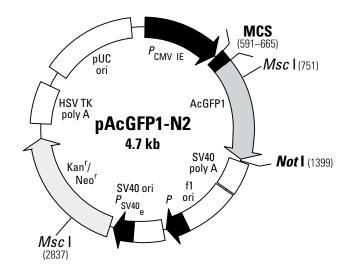
Cat. Nos. 632483 & 632485





**Restriction Map and Multiple Cloning Site (MCS) of pAcGFP1-N2.** Unique restriction sites are in bold. The *Not* I site follows the AcGFP1 stop codon. The *Nhe* I site cannot be used for fusions because it contains an in-frame stop codon. NOTE: The *Xba* I and *BcI* I sites are methylated in the DNA provided by Clontech Laboratories, Inc. If you wish to digest the vector with these enzymes, you will need to transform the vector into a dam– host and make fresh DNA.

### **Description:**

pAcGFP1-N2 encodes a Green Fluorescent Protein (GFP) from *Aequorea coerulescens*. (Excitation maximum = 475 nm; emission maximum = 505 nm.) The coding sequence of the AcGFP1 gene contains silent base changes, which correspond to human codon-usage preferences (1). The MCS in pAcGFP1-N2 is between the immediate early promoter of CMV ( $P_{\text{CMV IE}}$ ) and the AcGFP1 coding sequences. Genes cloned into the MCS will be expressed as fusions to the N-terminus of AcGFP1 if they are in the same reading frame as AcGFP1 and there are no intervening stop codons. SV40 polyadenylation signals downstream of the AcGFP1 gene direct proper processing of the 3' end of the AcGFP1 mRNA. The vector backbone also contains an SV40 origin for replication in mammalian cells expressing the SV40T antigen. A neomycin-resistance cassette (Neo¹)—consisting of the SV40 early promoter, the neomycin/kanamycin resistance gene ofTn5, and polyadenylation signals from the Herpes simplex virus thymidine kinase (HSVTK) gene—allows stably transfected eukaryotic cells to be selected using G418. A bacterial promoter upstream of the gene expresses kanamycin resistance in *E. coli*. The pAcGFP1-N2 backbone also provides a pUC origin of replication for propagation in *E. coli* and an f1 origin for single-stranded DNA production.

#### Use

Fusions to the N-terminus of AcGFP1 retain the fluorescent properties of the native protein allowing the localization of the fusion protein *in vivo*. The target gene should be cloned into pAcGFP1-N2 such that it is in frame with the AcGFP1 coding sequences and contains no intervening in-frame stop codons. The inserted gene should include the initiating ATG codon. The recombinant AcGFP1 vector can be transfected into mammalian cells using any standard transfection method. If required, stable transformants can be selected using G418 (2). pAcGFP1-N2 can also be used simply to express AcGFP1 in a cell line of interest (e.g., as a transfection marker).

(PR8Z2663; published 11 December 2008)



United States/Canada 800.662.2566 Asia Pacific

+1.650.919.7300

Europe

+33.(0)1.3904.6880

Japan +81.(0)77.543.6116

Clontech Laboratories, Inc. ATakara Bio Company 1290 Terra Bella Ave. Mountain View, CA 94043 Technical Support (US) E-mail: tech@clontech.com www.clontech.com pAcGFP1-N2 Vector Information

### **Location of Features:**

Human cytomegalovirus (CMV) immediate early promoter: 1–589

Enhancer region: 59-465; TATA box: 554-560

Transcription start point: 583

C→G mutation to remove Sac I site: 569

MCS: 591–665

 Aequorea coerulescens green fluorescent protein gene Start codon (ATG): 677–679; Stop codon: 1394–1396

Insertion of Val at position 2: 680–682
• SV40 early mRNA polyadenylation signal

Polyadenylation signals: 1549–1554 & 1578–1583; mRNA 3' ends: 1587 & 1599
• f1 single-strand DNA origin: 1646–2101 (Packages the noncoding strand of AcGFP1)

Bacterial promoter for expression of Kan<sup>r</sup> gene:

-35 region: 2163-2168; -10 region: 2186-2191

Transcription start point: 2198
• SV40 origin of replication: 2442–2577

SV40 early promoter

Enhancer (72-bp tandem repeats): 2275-2346 & 2347-2418

21-bp repeats: 2422-2442, 2443-2463 & 2465-2485

Early promoter element: 2498-2504

Major transcription start points: 2494, 2532, 2538 & 2543

Kanamycin/neomycin resistance gene

Neomycin phosphotransferase coding sequences: Start codon (ATG): 2626–2628; stop codon: 3418–3420

G→A mutation to remove Pst I site: 2808

C→A (Arg to Ser) mutation to remove *Bss*H II site: 3154

Herpes simplex virus (HSV) thymidine kinase (TK) polyadenylation signal

Polyadenylation signals: 3656-3661 & 3669-3674

pUC plasmid replication origin: 4005–4648

# Propagation in *E. coli*:

Suitable host strains: DH5α, HB101, and other general purpose strains. Single-stranded DNA production requires
a host containing an F plasmid such as JM109 or XL1-Blue.

Selectable marker: plasmid confers resistance to kanamycin (50 µg/ml) in E. coli hosts.

E. coli replication origin: pUC

Copy number: ≈500

Plasmid incompatibility group: pMB1/ColE1

### References:

1. Haas, J., et al. (1996) Curr. Biol. 6:315-324.

2. Gorman, C. (1985). In DNA cloning: A practical approach, vol. II. Ed. D.M. Glover. (IRL Press, Oxford, U.K.) pp. 143–190.

**Note:** The attached sequence file has been compiled from information in the sequence databases, published literature, and other sources, together with partial sequences obtained by Clontech Laboratories, Inc. This vector has not been completely sequenced.

## **Notice to Purchaser**

Clontech products are to be used for research purposes only. They may not be used for any other purpose, including, but not limited to, use in drugs, *in vitro* diagnostic purposes, therapeutics, or in humans. Clontech products may not be transferred to third parties, resold, modified for resale, or used to manufacture commercial products or to provide a service to third parties without written approval of Clontech Laboratories, Inc.

Not-For-Profit Entities: Orders may be placed in the normal manner by contacting your local representative or Clontech Customer Service at 650.919.7300. At its discretion, Clontech grants Not-For-Profit Entities a non-exclusive, royalty-free, personal, limited license to use this product for non-commercial life science research use only. Such license specifically excludes the right to sell or otherwise transfer this product, its components or derivatives thereof to third parties. No modifications to the protein coding sequence may be made without express written permission from Clontech. Any other use of this product requires a license from Clontech. For license information, please contact a licensing representative by phone at 650.919.7320 or by e-mail at licensing@clontech.com.

For-Profit Entities wishing to use this product are required to obtain a license from Clontech. For license information, please contact a licensing representative by phone at 650.919.7320 or by e-mail at licensing@clontech.com

Clontech, the Clontech logo and all other trademarks are the property of Clontech Laboratories, Inc., unless noted otherwise. Clontech is a Takara Bio Company. ©2008 Clontech Laboratories, Inc.