# Cogent<sup>™</sup> NGS Analysis Pipeline v3.0.19 (beta) Quick Start Guide

The following information is provided as a high-level introduction to the software, also referred to as CogentAP. For more detailed information, please see the <u>Cogent NGS Analysis Pipeline v2.0 User Manual</u>.

### Before you begin

- A. Supported operating systems
  - Linux: CentOS 8 or higher, RedHat 8 or higher, Ubuntu 18.04 or higher
- B. Hardware requirements
  - CPU: 24 cores
  - Memory: 64 GB RAM
  - Free disk space: at least 500 GB
- C. Additional dependencies
  - Internet connectivity on the server
  - Conda 24.4.0 or higher
  - Bash UNIX shell
- D. Required input files
  - FASTQ files generated by an Illumina® sequencing platform.
     NOTE: For a list of supported Takara Bio chemistries, please refer to our <u>bioinformatics</u> portal web page.
  - A well-list text file, Illumina sample sheet, or similar TDT/CSV format file

# **Confirm Conda version**

1. Verify conda is installed and meets or exceeds the required version by typing the following into a terminal window:

conda -V

If conda is successfully installed, it should return text with the version number.

Example:

conda 24.4.0

2. Verify the base conda environment can be activated by typing:

conda activate

If conda is successfully installed, the terminal will look similar to this:

```
$ conda activate
(base) $
```

Type the following command to return to the default Linux prompt.

conda deactivate

(base) \$ conda dectivate \$

- 3. Verify the install location of Miniconda3 is configured in the file .bash\_profile
  - a. For an individual user account, type: more ~/.bash\_profile
  - b. Confirm something similar to the following is showing in the file (all on one line): export PATH="/home/<USERNAME>/miniconda3/

bin:\$PATH"

where <USERNAME> is replaced by the username of the account that installed conda,

If no .bash\_profile file exists or the line isn't displaying, it will need to be manually created and populated.

# Installation

- 1. <u>Sign up</u> to download the installation package from our website.
- 2. Move or copy the ZIP file downloaded from Step 1 onto the Linux server into the directory location where you want to install.
- 3. Unzip the installation package (all on one line):

```
unzip Cogent_NGS_Analysis_Pipeline_v3.0.19.zip \
&& mv Cogent_NGS_Analysis_Pipeline_v3.0.19.zip \
CogentAP
```

 Run these commands sequentially to install the packages required to run CogentAP:
 cd CogentAP

bash CogentAP setup.sh install

5. After the conda environment packages install step completes, install the human genome (for the mouse genome, use mm10 rather than hg38):

bash CogentAP\_setup.sh genome\_install
hg38



## **Generation of raw FASTQ files**

- 1. Log in to a server that stores the run folder from Illumina sequencing and has the bcl2fastq program installed.
- 2. Change to a working folder where you want the raw FASTQ files to be located after being generated.
- 3. To convert BCL files to FASTQ files using bcl2fastq, *go to Step #4*. If using BCL Convert, *go to Step #5*.
- 4. Run bcl2fastq with the following syntax template:

```
bcl2fastq -R <RUN_FOLDER> -o <RUN_ID>
--no-lane-splitting --sample-sheet
$COGENT_AP_HOME/config/SampleSheet_dum
my.csv > <RUN_ID>.stdout 2 >
<RUN_ID>.stderr
```

#### where:

- <RUN\_FOLDER> is the path to the sequencing run folder
- <RUN\_ID> is the ID number automatically generated by the Illumina sequencer

The file SampleSheet\_dummy.csv is stored in the CogentAP config folder

#### Go to Step #6.

**NOTE:** See Section IV.D of the Cogent Analysis Pipeline v2.0 User Manual for how to set up the \$COGENT AP HOME variable.

5. Run BCL Convert with the following syntax template:

```
bcl-convert --bcl-input-directory
<RUN_FOLDER> --output-directory
<RUN_ID> --no-lane-splitting --sample-
sheet=DummySampleSheet >
<RUN ID>.stdout 2 > <RUN ID>.stderr
```

Templates for the DummySampleSheet for BCL Convert are stored in the CogentAP config folder.

#### Go to Step #6.

 Move the raw FASTQ files to your preferred storage location. They are typically generated in the <RUN\_ID> folder and named similar to:

```
Undetermined_S0_R1_001.fastq.gz
Undetermined_S0_R2_001.fastq.gz
```

# Running Cogent NGS Analysis Pipeline Software

• To demultiplex (demux)



```
<COGENT_AP_HOME>/cogent rna demux \

-f <FASTQ_R1> \

-p <FASTQ_R2> \

-b <WELL-LIST> \

-t <EXP_TYPE> \

-o <OUTPUT>
```

To analyze

```
<COGENT_AP_HOME>/cogent rna analyze \
-i <DEMUX_RESULT_DIR> \
-g <GENOME> \
-t <EXP_TYPE> \
-o <OUTPUT>
```

#### where:

- <COGENT\_AP\_HOME> is the path to the directory where CogentAP is installed
- <FASTQ\_R1> and <FASTQ\_R2> are the full paths to the FASTQ files generated by an Illumina sequencing platform.
- <WELL-LIST> is the full path to the ICELL8 system WellList, Illumina's sample sheet or TDT/CSV format file
- <EXP\_TYPE> is the experiment type used (e.g., icell8\_fla, refer to the CogentAP User Manual for more options)
- <OUTPUT> is a string; it will be the name of the output folder created by the analysis AND the prefix of all the results files
- <DEMUX\_RESULT\_DIR> is the full path of the demultiplex result directory you specified in demux command
- <GENOME> is a name of genome build (e.g., hg38)

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#### Takara Bio USA, Inc.

United States/Canada: +1.800.662.2566 • Asia Pacific: +1.650.919.7300 • Europe: +33.(0)1.3904.6880 • Japan: +81.(0)77.565.6999

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