





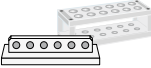


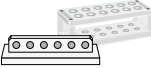


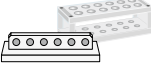


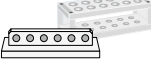


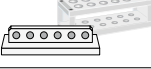

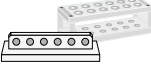


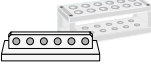


Protocol at a glance

Isolation of high molecular weight DNA

1 Prepare and lyse samples		25 μ L Proteinase K 600 μ L HM1 Mix or invert 56 °C, 30 - 150 min
2 Precipitate debris	 	11,000 x g, 1 min, 5 min Transfer up to 500 μ L to a fresh 2 mL tube
3 Digest RNA		20 μ L RNase Mix or invert 15 min, RT
4 Bind DNA to NucleoMag® B-Beads	  	25 μ L NucleoMag® B-Beads 400 μ L HM2 Mix by shaking for 5 min at 1000 rpm (Optional: Mix by pipetting up and down using wide-bore tips) Remove supernatant after 5 min separation
5 Wash with HM3	  	Remove tubes from NucleoMag® SEP Mini 900 μ L HM3 Mix by shaking for 2 min, 1000 rpm (Optional: Mix by pipetting up and down using wide-bore tips) Remove supernatant after 2 min separation
6 Wash with HM4	  	Remove tubes from NucleoMag® SEP Mini 900 μ L HM4 Mix by shaking for 2 min, 1000 rpm (Optional: Mix by pipetting up and down using wide-bore tips) Remove supernatant after 2 min separation
7 1st Wash with 70 % EtOH	  	Remove tubes from NucleoMag® SEP Mini 900 μ L 70 % EtOH Mix by shaking for 2 min, 1000 rpm (Optional: Mix by pipetting up and down using wide-bore tips) Remove supernatant after 2 min separation
8 2nd wash with 70 % EtOH	  	Remove tubes from NucleoMag® SEP Mini 900 μ L 70 % EtOH Mix by shaking for 2 min, 1000 rpm (Optional: Mix by pipetting up and down using wide-bore tips) Remove supernatant after 2 min separation
9 Rinse the beads	 	Leave tubes on NucleoMag® SEP Mini 600 μ L HM5 Incubate for 30–60 s <i>Note: Do not resuspend the beads in Buffer HM5!</i> Remove supernatant
10 Elute DNA	  	Remove tubes from NucleoMag® SEP Mini 100 μ L HM6 Shake 5 min at RT 1000 rpm (Optional: Mix by pipetting up and down) using wide-bore tips Separate 2 min and transfer DNA into elution plate / tubes



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