



SMART-Seq® DE3 Demultiplexer User Guide

I. Introduction

The SMART-Seq DE3 Demultiplexer is bioinformatics software for use only with sequencing data generated from [SMART-Seq mRNA 3' DE](#). This software uses in-line indexes incorporated in paired reads to demultiplex your samples and generate an output file with sorted read data. These in-line indexes are hard-coded in the software.

More specifically, SMART-Seq mRNA 3' DE generates a library in which unique in-line indexes are added to the cDNA to serve as cell barcodes and enable sample pooling prior to library preparation and sequencing. The library produces paired reads where the in-line index is encoded in the first six nucleotides of read 2 and the sequence of interest is contained in paired read 1. For more information on SMART-Seq mRNA 3' DE, please visit the [product page](#).

NOTE: If you know you have Java 1.7 or later installed and are only looking for the command arguments, please see Section IV.B.

II. Before You Begin

A. Input Files

Input files are paired reads, where read 1 contains a sequence of interest and read 2 contains a corresponding barcode sequence. SMART-Seq DE3 Demultiplexer works with either gzipped files (.gz extension) or uncompressed files (commonly with .fq or .fastq extensions).

B. Verifying Java and Confirming Operating System

SMART-Seq DE3 Demultiplexer is written in Java and is compatible with any operating system where Java 1.7 or newer is installed. We have tested SMART-Seq DE3 Demultiplexer on Linux (Ubuntu 16.40), Windows (7 and 10) and Mac (OS X El Capitan).

After opening a terminal window, verify that you have the correct version of Java installed by typing the following command into your terminal window (see section IV.A for instructions on opening a terminal window):

```
java -version
```

```
$ java -version
java version "1.8.0_91"
Java(TM) SE Runtime Environment (build 1.8.0_91-b14)
Java HotSpot(TM) 64-Bit Server VM (build 25.91-b14, mixed mode)
$
```

Figure 1. After typing `java -version` into your terminal, you should see a version number displayed. Verify that the first two numbers are 1.7 or higher.

C. Example Data

We have provided sample data in the zip archive that you can use to confirm operation of the demultiplexing software. The paired-end sample data contains a pooled library of 2,500 reads and all 12 in-line indexes. This representative dataset was acquired with a library synthesized with SMART-Seq mRNA 3' DE and sequenced on an Illumina® MiSeq® instrument. The data are contained in two files named:

- `Example_Data_R1_.fastq.gz` (from read 1)
- `Example_Data_R2_.fastq.gz` (from read 2)

All examples below refer to this sample data.

III. Installing SMART-Seq DE3 Demultiplexer

The software is available to download as a ZIP file from

<https://www.takarabio.com/products/next-generation-sequencing/bioinformatics-tools/smart-seq-de3-demultiplexer>.

Unzip the archive in a convenient directory. The software is packaged as an executable Java JAR file. No additional steps are necessary to install the software.

IV. Running SMART-Seq DE3 Demultiplexer

SMART-Seq DE3 Demultiplexer is written as a command-line-interface tool and is executed from within a terminal window.

A. Opening a Terminal Window

- **Mac:** The Terminal application is typically found under **Applications > Utilities > Terminal**
- **Linux (Ubuntu):** Use the keyboard shortcut **[Ctrl][Alt][T]**.

Alternatively, you can find the terminal by opening the [Dash] (upper left on most desktops), typing `terminal` and selecting the Terminal application.

- **Windows:** Click the [Start] button and type `cmd` in the run box.

Alternatively, select "Command Prompt" from **Start > Program Files > Accessories**.

B. Running the Demultiplexing Command

The command to demultiplex your sequencing run is (all in one command):

```
java -jar \  
/path/to/SMART_Seq_DE3_Demultiplexer.jar \  
/path/to/read1 \  
/path/to/read2
```

NOTE: on Mac and Linux systems ~/ is not expanded automatically by Java.

- Use the full path, i.e.,
/home/tbusa/SMART_Seq_DE3_Demultiplexer/Example_Data_R1_.fastq.gz
- Do not use
~/SMART_Seq_DE3_Demultiplexer/Example_Data_R1_.fastq.gz

Table 1 provides a brief explanation of the command:

Table 1. SMART_Seq_DE3_Demultiplexer command breakdown.

Column 1	Column 2
java -jar	System command to execute a Java jar file
/path/to/SMART_Seq_DE3_Demultiplexer.jar	Full system path to the SMART-Seq DE3 Demultiplexer jar file
/path/to/read1	Full system path to read 1 FASTQ file
/path/to/read2	Full system path to read 2 FASTQ file

As an alternative to typing the demultiplex command, you can drag and drop files into the command window. If you'd like to avoid typing out the full file paths, you can drag a file onto the terminal window to automatically add each file path to the end of the current command line. In order to use this method to fill out the full command, use the following steps:

1. Open the terminal window and the folder containing the SMART_Seq_DE3_Demultiplexer.jar file and your read data.
2. Type: `java -jar`
3. Press **[Space bar]**.
4. Drag and drop the SMART_Seq_DE3_Demultiplexer.jar file into the terminal window (you should now see a full path to the executable file).
5. Press **[Space bar]**.
6. Drag and drop the read 1 file into the terminal window (you should see the full path for the FASTQ file).
7. Press **[Space bar]**.
8. Drag and drop the read 2 file into the terminal window (you should see the full path for the FASTQ file).

C. Example Command

Here, we provide an example command run on Linux with the zip archive extracted into our home folder (/home/tbusa/). The following command will demultiplex the example data included in the archive.

NOTE: The following command is a single line.

```
java -jar \
/home/tbusa/SMART_Seq_DE3_Demultiplexer/SMART_Seq_DE3_Demultiplexer.jar \
/home/tbusa/SMART_Seq_DE3_Demultiplexer/Example_Data_R1_.fastq.gz \
/home/tbusa/SMART_Seq_DE3_Demultiplexer/Example_Data_R2_.fastq.gz
```

V. Examining the Demultiplexing Command Output

A. Output Files

The demultiplexing command always produces 26 files, even if no sequences are placed in a file (e.g. if you are analyzing data from 6 pooled samples, there will still be 26 files produced by the software). The original data files are maintained in their original directory. Three kinds of files are produced by the demultiplexing command:

- **Read 1:** There are 12 “read 1” output files that are named with the in-line index added to the beginning of the original file name and the extension `.fastq` added to the end (e.g., `IL01_Example_Data_R1_.fastq.gz.fastq`). These files now contain sequencing reads that are usable for downstream analyses.
- **Read 2:** There are 12 “read 2” output files that are named similarly to the read 1 output files (e.g., `IL01_Example_Data_R2_.fastq.gz.fastq`). These files are provided to the user to verify that the demultiplexing command performed as expected. For instance, one might verify that the sequences in the `IL12_*` file all start with `CTTGTA`; this sequence corresponds to in-line index 12.
- **Undetermined files:** There are two `UNDETERMINED` files, one associated with read 1 and one associated with read 2. When the software cannot match the first six nucleotides in read 2 to one of the twelve expected barcodes, it places those reads in the `UNDETERMINED_*` file-pair.

Name	Size	Type	Modified
Example_Data_R1_fastq.gz	229.9 kB	Archive	May 24
Example_Data_R2_fastq.gz	21.8 kB	Archive	May 24
IL01_Example_Data_R1_fastq.gz.fastq	67.6 kB	Text	09:48
IL01_Example_Data_R2_fastq.gz.fastq	24.3 kB	Text	09:48
IL02_Example_Data_R1_fastq.gz.fastq	61.8 kB	Text	09:48
IL02_Example_Data_R2_fastq.gz.fastq	22.2 kB	Text	09:48
IL03_Example_Data_R1_fastq.gz.fastq	65.5 kB	Text	09:48
IL03_Example_Data_R2_fastq.gz.fastq	23.5 kB	Text	09:48
IL04_Example_Data_R1_fastq.gz.fastq	62.4 kB	Text	09:48
IL04_Example_Data_R2_fastq.gz.fastq	22.4 kB	Text	09:48
IL05_Example_Data_R1_fastq.gz.fastq	67.9 kB	Text	09:48
IL05_Example_Data_R2_fastq.gz.fastq	24.4 kB	Text	09:48
IL06_Example_Data_R1_fastq.gz.fastq	55.1 kB	Text	09:48
IL06_Example_Data_R2_fastq.gz.fastq	19.8 kB	Text	09:48
IL07_Example_Data_R1_fastq.gz.fastq	67.6 kB	Text	09:48
IL07_Example_Data_R2_fastq.gz.fastq	24.3 kB	Text	09:48
IL08_Example_Data_R1_fastq.gz.fastq	58.1 kB	Text	09:48
IL08_Example_Data_R2_fastq.gz.fastq	20.9 kB	Text	09:48
IL09_Example_Data_R1_fastq.gz.fastq	59.7 kB	Text	09:48
IL09_Example_Data_R2_fastq.gz.fastq	21.4 kB	Text	09:48
IL10_Example_Data_R1_fastq.gz.fastq	68.2 kB	Text	09:48
IL10_Example_Data_R2_fastq.gz.fastq	24.5 kB	Text	09:48
IL11_Example_Data_R1_fastq.gz.fastq	65.5 kB	Text	09:48
IL11_Example_Data_R2_fastq.gz.fastq	23.5 kB	Text	09:48
IL12_Example_Data_R1_fastq.gz.fastq	45.9 kB	Text	09:48
IL12_Example_Data_R2_fastq.gz.fastq	16.5 kB	Text	09:48
SMART_Seq_DE3_Demultiplexer.jar	18.9 kB	Archive	May 24
UNDETERMINED_Example_Data_R1_fastq.gz.fastq	19.6 kB	Text	09:48
UNDETERMINED_Example_Data_R2_fastq.gz.fastq	7.0 kB	Text	09:48

Figure 2. This image displays all of the output files created after successfully running the demultiplexing software on the provided sample data. Twenty-six new files are generated and added to the directory with the original data file.

B. Command Summary

A summary of the demultiplexing results is written to the screen as a table with three columns:

- In-line index
- Output file path
- Number of sequencing reads that correspond to each index

The table is sorted by the name of the in-line index, with the UNDETERMINED sequences listed at the end. In the example shown in Figure 3 (next page), notice that each file is prefixed with the in-line index sequence and appended with the FASTQ extension, as discussed in Section V.A above.

```

$ java -jar /home/clontech/SMART_Seq_DE3_Demultiplexer/SMART_Seq_DE3_Demultiplexer.jar /home/clontech/SMART_Seq_DE3_Demultiplexer/Example_Data_R1.fastq.gz /home/clontech/SMART_Seq_DE3_Demultiplexer/Example_Data_R2.fastq.gz
Summary:
Barcode File Number_of_Records
CTAGCT /home/clontech/SMART_Seq_DE3_Demultiplexer/IL01_Example_Data_R1.fastq.gz.fastq 221
CGATGT /home/clontech/SMART_Seq_DE3_Demultiplexer/IL02_Example_Data_R1.fastq.gz.fastq 202
TTAGGC /home/clontech/SMART_Seq_DE3_Demultiplexer/IL03_Example_Data_R1.fastq.gz.fastq 214
TGACCA /home/clontech/SMART_Seq_DE3_Demultiplexer/IL04_Example_Data_R1.fastq.gz.fastq 204
ACAGTG /home/clontech/SMART_Seq_DE3_Demultiplexer/IL05_Example_Data_R1.fastq.gz.fastq 222
TATAAT /home/clontech/SMART_Seq_DE3_Demultiplexer/IL06_Example_Data_R1.fastq.gz.fastq 180
CAGATC /home/clontech/SMART_Seq_DE3_Demultiplexer/IL07_Example_Data_R1.fastq.gz.fastq 221
ACTTGA /home/clontech/SMART_Seq_DE3_Demultiplexer/IL08_Example_Data_R1.fastq.gz.fastq 190
GATCAG /home/clontech/SMART_Seq_DE3_Demultiplexer/IL09_Example_Data_R1.fastq.gz.fastq 195
TAGCTT /home/clontech/SMART_Seq_DE3_Demultiplexer/IL10_Example_Data_R1.fastq.gz.fastq 223
GGCTAC /home/clontech/SMART_Seq_DF3_Demultiplexer/IL11_Example_Data_R1.fastq.gz.fastq 214
CTTGTA /home/clontech/SMART_Seq_DE3_Demultiplexer/IL12_Example_Data_R1.fastq.gz.fastq 150
UNDETERMINED /home/clontech/SMART_Seq_DE3_Demultiplexer/UNDETERMINED_Example_Data_R1.fastq.gz.fastq 64
$

```

Figure 3. After running the command, the software will write a summary to the terminal window. The summary will display each in-line index (or UNDETERMINED for sequences that can't be matched), the output file for read 1 associated with each inline index, and the number of sequences that were sorted into each file.

VI. Technical Support

For any technical support regarding this protocol, please visit: takarabio.com/support.

Products

Cat. #	Product	Size
635040	SMART-Seq mRNA 3' DE	96 Rxns
635041		192 Rxns

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Customer Service/Ordering	Technical Support
tel: 800.662.2566 (toll-free)	tel: 800.662.2566 (toll-free)
fax: 800.424.1350 (toll-free)	fax: 800.424.1350 (toll-free)
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e-mail: ordersUS@takarabio.com	e-mail: technical_support@takarabio.com

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