# In-Fusion® Ready Vector Cloning Protocol-At-A-Glance (PT3865-2)

This protocol is provided for cloning PCR fragments into In-Fusion Ready vectors using In-Fusion HD Cloning Plus (Cat. Nos. 638909, 638910, 638911 and 638920).

## A. PCR Amplification for In-Fusion Ready Cloning

The sense and antisense primers that will be used to amplify the gene of interest via PCR must contain a specific 15 nucleotide sequence 5' to the sequence of the gene of interest, as shown below.

Sense primer: 5'-AAGGCCTCTGTCGAC followed by sequence of amplification target-3' Antisense primer: 5'-AGAATTCGCAAGCTT followed by sequence of amplification target-3'

**NOTE:** The PCR product obtained from amplification using these primers is also ready for In-Fusion cloning into all other Clontech prelinearized In-Fusion Ready vectors if the gene of interest contains an initiating ATG. For additional information regarding In-Fusion cloning, see the In-Fusion HD Cloning Kit User Manual.

## **B.** Cloning Procedure

- 1. Combine the following in an eppendorf tube:
  - 1 μl Linearized In-Fusion Ready Vector (100 ng/μl)
  - \_\_ μl (50 ng) Purified PCR product
  - 2 μl 5x In-Fusion HD Enzyme Premix
  - μl Sterile H<sub>2</sub>O

10 µl Total Volume

- 2. Incubate the reactions for 15 min at 50°C, then place on ice.
- 3. Proceed with transformation (Section C). If you cannot transform cells immediately, store cloning reactions at -20°C until you are ready.

### C. Transformation

- 1. Transform Stellar<sup>TM</sup> competent cells with 2.5 µl of reaction mixture.
  - a. Thaw one vial of frozen competent cells on ice. Tap tube gently to ensure that the cells are suspended.
  - **b.** Add 2.5 μl of the reaction mixture to the cells, mix gently to ensure even distribution of the DNA solution. Leave the tube on ice for 30 min.

### Do not add more than 5 µl of reaction to 50 µl of competent cells.

- c. Heat shock the cells in a water bath at 42°C for 45 sec, and then place them directly on ice for 1 min.
- 2. Add 450 µl of SOC medium to the cells and then incubate at 37°C for 60 min while shaking at 250 rpm.
- 3. Take 1/100 of the cells (5  $\mu$ l) from each transformation. Bring the volume up to  $100~\mu$ l with SOC medium, and plate by spreading different volumes on LB plates containing the appropriate antibiotic. Spread the remaining cells from each transformation on additional plates. Incubate all plates at  $37^{\circ}$ C overnight.
- 4. The next day, pick colonies from each experimental plate and isolate plasmid DNA using a standard method of your choice.

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