



**Restriction Map and Multiple Cloning Site (MCS) of pAcGFP1-C3.** Restriction sites shown in bold are unique. The *Msc I* site is not unique. NOTE: The *Xba I* and *Bcl I* sites are methylated in the DNA provided by Clontech Laboratories, Inc. If you wish to digest the vector with these enzymes, you will need to transform the vector into a dam<sup>-</sup> host and make fresh DNA.

### Description:

pAcGFP1-C3 encodes a Green Fluorescent Protein (GFP) from *Aequorea coerulescens*. (Excitation maximum = 475 nm; emission maximum = 505 nm.) Sequences flanking AcGFP1 have been converted to a Kozak consensus translation initiation site (1) to further increase the translation efficiency in eukaryotic cells. The MCS in pAcGFP1-C3 is downstream of the AcGFP1 coding region, allowing the construction of a C-terminal fusion protein with AcGFP1 when genes are cloned in the same reading frame as AcGFP1 and there are no intervening stop codons. SV40 polyadenylation signals downstream of the AcGFP1 gene direct proper processing of the 3' end of the AcGFP1 mRNA. The vector backbone also contains an SV40 origin for replication in mammalian cells expressing the SV40 T-antigen. A neomycin resistance cassette (Neo<sup>r</sup>)—consisting of the SV40 early promoter, the neomycin/kanamycin resistance gene of Tn5, and polyadenylation signals from the Herpes simplex virus thymidine kinase (HSV TK) gene—allows stably transfected eukaryotic cells to be selected using G418. A bacterial promoter upstream of the gene expresses kanamycin resistance in *E. coli*. The pAcGFP1-C3 backbone also provides a pUC origin of replication for propagation in *E. coli* and an f1 origin for single-stranded DNA production.

### Use:

Fusions to the C-terminus of AcGFP1 retain the fluorescent properties of the native protein, allowing the localization of the fusion protein *in vivo*. The target gene should be cloned into pAcGFP1-C3 such that it is in frame with the AcGFP1 coding sequences and contains no intervening in-frame stop codons. The recombinant AcGFP1 vector can be transfected into mammalian cells using any standard transfection method. If required, stable transformants can be selected using G418 (2). pAcGFP1-C3 can also be used simply to express AcGFP1 in a cell line of interest (e.g., as a transfection marker).

(PR651714; published 2 May 2006)



**Clontech**

United States/Canada  
800.662.2566

Asia Pacific  
+1.650.919.7300

Europe  
+33.(0)1.3904.6880

Japan  
+81.(0)77.543.6116

Clontech Laboratories, Inc.  
A Takara Bio Company  
1290 Terra Bella Ave.  
Mountain View, CA 94043  
Technical Support (US)  
E-mail: tech@clontech.com  
www.clontech.com

**Location of Features:**

- Human cytomegalovirus (CMV) immediate early promoter: 1–589  
Enhancer region: 59–465; TATA box: 554–560  
Transcription start point: 583  
C→G mutation to remove *Sac* I site: 569
- *Aequorea coerulescens* green fluorescent protein gene  
Kozak consensus translation initiation site: 606–616  
Start codon (ATG): 613–615; Stop codon: 1408–1410  
Insertion of Val at position 2: 616–618  
Last amino acid: 1327–1329
- MCS: 1328–1413
- SV40 early mRNA polyadenylation signal  
Polyadenylation signals: 1546–1551 & 1575–1580; mRNA 3' ends: 1584 & 1596
- f1 single-strand DNA origin: 1643–2098 (Packages the noncoding strand of AcGFP1)
- Bacterial promoter for expression of Kan<sup>r</sup> gene  
–35 region: 2160–2165; –10 region: 2183–2188  
Transcription start point: 2195
- SV40 origin of replication: 2439–2574
- SV40 early promoter  
Enhancer (72-bp tandem repeats): 2272–2343 & 2344–2415  
21-bp repeats: 2419–2439, 2440–2460 & 2462–2482  
Early promoter element: 2495–2501  
Major transcription start points: 2491, 2529, 2535 & 2540
- Kanamycin/neomycin resistance gene  
Neomycin phosphotransferase coding sequences:  
Start codon (ATG): 2623–2625; stop codon: 3415–3417  
G→A mutation to remove *Pst* I site: 2805  
C→A (Arg to Ser) mutation to remove *Bss*H II site: 3151
- Herpes simplex virus (HSV) thymidine kinase (TK) polyadenylation signal  
Polyadenylation signals: 3653–3658 & 3666–3671
- pUC plasmid replication origin: 4002–4645

**Propagation in *E. coli*:**

- Suitable host strains: DH5 $\alpha$ , HB101, and other general purpose strains. Single-stranded DNA production requires a host containing an F plasmid such as JM109 or XL1-Blue.
- Selectable marker: plasmid confers resistance to kanamycin (50  $\mu$ g/ml) in *E. coli* hosts.
- *E. coli* replication origin: pUC
- Copy number:  $\approx$ 500
- Plasmid incompatibility group: pMB1/ColE1

**References:**

1. Kozak, M. (1987) *Nucleic Acids Res.* **15**:8125–8148.
2. Gorman, C. (1985) In *DNA Cloning: A Practical Approach, Vol. II*, Ed. Glover, D. M. (IRL Press, Oxford, UK) pp. 143–190.

**Note:** The attached sequence file has been compiled from information in the sequence databases, published literature, and other sources, together with partial sequences obtained by Clontech Laboratories, Inc. This vector has not been completely sequenced.

**Notice to Purchaser**

This product is intended to be used for research purposes only. It is not to be used for drug or diagnostic purposes nor is it intended for human use. Clontech products may not be resold, modified for resale, or used to manufacture commercial products without written approval of Clontech Laboratories, Inc.

Not-For-Profit Entities: Orders may be placed in the normal manner by contacting your local representative or Clontech Customer Service at 650.919.7300. At its discretion, Clontech grants Not-For-Profit Entities a non-exclusive, personal, limited license to use this product for non-commercial life science research use only. Such license specifically excludes the right to sell or otherwise transfer this product, its components or derivatives thereof to third parties. No modifications to the protein coding sequence may be made without express written permission from Clontech. Any other use of this product requires a license from Clontech. For license information, please contact a licensing representative by phone at 650.919.7320 or by e-mail at [licensing@clontech.com](mailto:licensing@clontech.com).

For-Profit Entities wishing to use this product are required to obtain a license from Clontech. For license information, please contact a licensing representative by phone at 650.919.7320 or by e-mail at [licensing@clontech.com](mailto:licensing@clontech.com).

Clontech, Clontech logo and all other trademarks are the property of Clontech Laboratories, Inc. Clontech is a Takara Bio Company. ©2005