pDsRed2-C1 Vector Information



Restriction Map and Multiple Cloning Site (MCS) of pDsRed2-C1. Unique restriction sites are in bold.

Description

pDsRed2-C1 encodes DsRed2, a DsRed variant that has been engineered for faster maturation and lower non-specific aggregation. Derived from the Discosoma sp. red fluorescent protein (drFP583; 1), DsRed2, like its progenitor DsRed1, contains a series of silent base-pair changes that correspond to human codon-usage preferences for high expression in mammalian cells (2). In addition to these changes, DsRed2 contains six amino acid substitutions: V105A, I161T, and S197A, which result in the more rapid appearance of red fluorescence in transfected cell lines; and R2A, K5E, and K9T, which prevent the protein from aggregating. (DsRed2 may, however, form the same tetrameric structure as DsRed1 [3].) In mammalian cell cultures when DsRed2 is expressed constitutively, red-emitting cells can be detected by fluorescence microscopy within 24 hours of transfection. Large insoluble aggregates of protein, often observed in bacterial and mammalian cell systems expressing DsRed1, are dramatically reduced in organisms expressing DsRed2. The faster-maturing, more soluble red fluorescent protein is also well tolerated by host cells; mammalian cell cultures transfected with DsRed2 show no obvious signs of reduced viability -- in those cell lines tested, cells expressing DsRed2 display the same morphology (e.g., adherence, light-refraction) and growth characteristics as nontransfected controls.

The multiple cloning site (MCS) in pDsRed2-C1 is positioned between the DsRed2 coding sequence and the SV40 polyadenylation signal (SV40 poly A). Genes cloned into the MCS will be expressed as fusions to the C-terminus of DsRed2 if they are in the same reading frame as DsRed2 and there are no intervening stop codons. A Kozak consensus translation initiation site upstream of DsRed2 increases the translation efficiency in eukaryotic cells (4). SV40 poly A signals downstream of the MCS direct proper processing of the 3' end of mRNA transcripts. The vector backbone also contains an SV40 origin for replication in mammalian cells expressing the SV40 T-antigen, a pUC origin of replication for propagation in *E. coli*, and an f1 origin for single-stranded DNA production. A neomycin resistance cassette (Neo^r), consisting of the SV40 early promoter, the neomycin/kanamycin resistance gene of Tn5, and polyadenylation signals from the Herpes simplex virus thymidine kinase (HSVTK) gene, allows stably transfected eukaryotic cells to be selected using G418. A bacterial promoter upstream of this cassette expresses kanamycin resistance in *E. coli*.

(PR063555; published 30 June 2010)



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Use

pDsRed2-C1 can be used to construct fusions to the C-terminus of DsRed2. If a fusion construct retains the fluorescent properties of the native DsRed2 protein, its expression can be monitored by flow cytometry and its localization *in vivo* can be determined by fluorescence microscopy. The target gene should be cloned into pDsRed2-C1 so that it is in frame with the DsRed2 coding sequences, with no intervening in-frame stop codons. The recombinant DsRed2 vector can be transfected into mammalian cells using any standard transfection method. If required, stable transformants can be selected using G418 (5). pDsRed2-C1 can also be used as a cotransfection marker; the unmodified vector will express DsRed2.

Location of features

- Human cytomegalovirus (CMV) immediate early promoter: 1–589 Enhancer region: 59–465; TATA box: 554–560 Transcription start point: 583
 Construction of the promote and point: 560
 - $C \rightarrow G$ mutation to remove *Sac* I site: 569
- Discosoma sp. human codon-optimized Red Fluorescent Protein (DsRed2) gene Kozak consensus translation initiation site: 606–616 Start codon (ATG): 613–615; Stop codon: 1357–1359 Last amino acid in DsRed2: 1285–1287
- MCS: 1288-1344
- SV40 early mRNA polyadenylation signal Polyadenylation signals: 1498–1503 & 1527–1532; mRNA 3' ends: 1536 & 1648
- f1 single-strand DNA origin: 1595–2050 (Packages the noncoding strand of DsRed2.)
- Bacterial promoter for expression of Kan^r gene –35 region: 2112–2117; –10 region: 2135–2140 Transcription start point: 2147
- SV40 origin of replication: 2391–2526
- SV40 early promoter Enhancer (72-bp tandem repeats): 2224–2295 & 2296–2367 21-bp repeats: 2371–2991, 2392–2412 & 2414–2431 Early promoter element: 2447–2453 Major transcription start points: 2443, 2481, 2487 & 2492
- Kanamycin/neomycin resistance gene Neomycin phosphotransferase coding sequences: Start codon (ATG): 2576–2578; stop codon: 3368–3370 G→A mutation to remove *Pst* I site: 2758 C→A (Arg to Ser) mutation to remove *Bss*H II site: 3104
- Herpes simplex virus (HSV) thymidine kinase (TK) polyadenylation signal Polyadenylation signals: 3606–3611 & 3619–3624
- pUC plasmid replication origin: 3955-4598

Sequencing primer locations

- DsRed1-N Sequencing Primer (Cat. No. 632387; 5'-GTACTGGAACTGGGGGGGACAG-3'): 813-792
- DsRed1-C Sequencing Primer (Cat. No. 632388; 5'-AGCTGGACATCACCTCCCACAACG-3'): 1205–1228

Propagation in *E. coli*

- Suitable host strains: DH5α, HB101, and other general purpose strains. Single-stranded DNA production requires a host containing an F plasmid such as JM109 or XL1-Blue.
- Selectable marker: plasmid confers resistance to kanamycin (50 µg/ml) to *E. coli* hosts.
- E. coli replication origin: pUC
- Copy number: ≈500
- Plasmid incompatibility group: pMB1/Col E1

Red Fluorescent Protein (DsRed2)

• Excitation/Emission Maxima: 558 nm / 583 nm

References:

- 1. Matz, M. V., et al. (1999) Nature Biotech. 17:969–973.
- 2. Haas, J., et al. (1996) Curr. Biol. 6:315-324.
- 3. Yarbrough, D., et al. (2001) Proc. Natl. Acad. Sci. USA 98:462-467.
- 4. Kozak, M. (1987) Nucleic Acids Res. 15:8125–8148.
- 5. Gorman, C. (1985) In DNA cloning: A Practical Approach, Vol. II. Ed. D. M. Glover. (IRL Press, Oxford, U.K.), pp. 143–190.

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The RCFP's (including DsRedExpress and DsRedExpress2) are covered by one or more of the following U.S. Patent Nos. 7,166,444; 7,157,565; 7,217,789; 7,338,784; 7,338,783; 7,537,915 6,969,597, 7,150,979 and 7,442,522.

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